

# PCSK9 immunization using nanoliposomes: preventive efficacy against hypercholesterolemia and atherosclerosis

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## Abstract

**Introduction:** The aim of the study was to study a nanoliposomal anti-PCSK9 vaccine as a novel approach for cholesterol lowering via PCSK9 inhibition.

**Material and methods:** An immunogenic peptide construct termed immunogenic fused PCSK9-tetanus (IFPT) was displayed on the surface of liposome nanoparticles (L-IFPT) and mixed into alum adjuvant (L-IFPTA+). The manufactured vaccine formulations IFPT, L-IFPT, L-IFPTA+, IFPTA+, and free nanoliposomes were subcutaneously injected four times with bi-weekly intervals in C57BL/6 mice on a severe atherogenic protocol.

**Results:** Among the formulations, L-IFPTA+ vaccine was found to elicit the highest IgG response against PCSK9 peptide. The induced PCSK9 antibodies inhibited PCSK9-LDLR interaction through binding to PCSK9 in vaccinated mice. Liver low-density lipoprotein receptor (LDLR) protein was increased in vaccinated mice. L-IFPTA+, L-IFPT and IFPTA+ vaccines reduced total cholesterol by up to  $-38.13 \pm 3.8\%$  ( $p = 0.006$ ),  $-23 \pm 4.1\%$  ( $p = 0.027$ ) and  $-19.12 \pm 3\%$  ( $p = 0.038$ ), and low-density lipoprotein cholesterol (LDL-C) by up to  $-57 \pm 7.7\%$  ( $p = 0.0003$ ),  $-41.67 \pm 4.2\%$  ( $p = 0.03$ ) and  $-36.11 \pm 5\%$  ( $p = 0.02$ ) in hypercholesterolemic mice, respectively, versus control mice after 8 weeks. Long-term assessment indicated that the vaccine formulations could stimulate a long-lasting humoral immune response against PCSK9 peptide, which was associated with a marked reduction of total cholesterol in L-IFPTA+, L-IFPT and IFPTA+ vaccine groups by up to  $-82.5 \pm 7.3\%$  ( $p = 0.002$ ),  $-70.54 \pm 6.2\%$  ( $p = 0.013$ ) and  $-72.02 \pm 8.7\%$  ( $p = 0.004$ ), respectively, and LDL-C by up to  $-88.14 \pm 5.6\%$  ( $p = 0.002$ ),  $-55.92 \pm 8.3\%$  ( $p = 0.003$ ) and  $54.81 \pm 9.3\%$  ( $p = 0.003$ ), respectively, versus the pre-vaccination time point

adjusted to the control group. Anti-inflammatory Th2 cells and IL-4 cytokine were considerably increased in splenocytes of vaccinated mice.

**Conclusions:** L-IFPTA+ vaccine can induce long-lasting, functional and safe PCSK9-specific antibodies in hypercholesterolemic C57BL/6 mice, providing a long-term protective impact on dyslipidemia and atherosclerosis.

**Key words:** PCSK9, immunization, atherosclerosis, dyslipidemia.

## Introduction

Low-density lipoprotein (LDL) cholesterol (LDL-C) lowering is known to be a safe and effective therapeutic approach for atherosclerotic cardiovascular disease [1]. Patients achieving extremely low LDL levels have lower incidence of cardiovascular events as evidenced by several outcome trials [2–5]. The clinical evidence demonstrates that extremely low plasma levels of LDL-C directed by elevated LDLR activity are not correlated with any considerable adverse effects [1].

LDL-C is mainly cleared from the bloodstream by the hepatic LDLR under the control of proprotein convertase subtilisin/kexin 9 (PCSK9). This enzyme plays a crucial role in the metabolism of plasma LDL-C, and PCSK9 inhibitors potently decrease elevated LDL-C concentrations in patients with familial hypercholesterolemia (FH) and/or cardiovascular disease (CVD) who did not achieve optimal LDL-C levels in spite of being on standard-of-care treatments, such as high-intensity statin therapy [6–8]. PCSK9 inhibitors are also known to reduce the coronary atherosclerosis burden. Imaging studies performed in the GLAGOV trial showed that adding evolocumab on top of statins augmented the impact of statins in reducing atheroma volume [9].

In recent years, several PCSK9-inhibiting methods have been developed, including gene silencing using RNA interference strategies, and inhibition of PCSK9 binding to LDLR using monoclonal antibodies (mAbs) or small peptides. The use of mAbs is currently the only approved approach for the inhibition of PCSK9-LDLR interaction [10–13]. However, due to their relatively limited *in vivo* half-lives, long-term administration of mAbs has drawbacks in terms of cost and compliance. Such limitations could be circumvented by active vaccination. More recently, some preclinical studies have shown the effectiveness of peptide-based active vaccination against PCSK9 in mouse models of hypercholesterolemia [14–17]. The main limitation of peptide antigens is their low immunogenicity. Adjuvant systems are commonly used as an effective strategy in vaccine development for improving the immunogenicity of antigenic peptides. Some adjuvants, such as virus-like particles [17], DNA [16], and keyhole limpet hemocyanin (KLH) [14, 15], have been used to intensify the immunogenicity of peptide-based antiPCSK9

vaccines; however, their safety and efficiency in human are still unclear.

Tetanus toxin is an approved adjuvant carrier used in human vaccines [18], and we utilized it as a peptide adjuvant covalently linked to PCSK9 antigen to enhance the vaccine immunogenicity. On the other hand, displaying the self-antigens in a repetitive and highly dense structure is an effective strategy for provoking potent antibody responses against self-antigens [19, 20]. Liposome are biocompatible and biodegradable bilayer vesicles that as adjuvant delivery systems have widely been employed in vaccine compositions. These spherical particles are feasible carriers for displaying antigens in peptide-based vaccines [21]. We previously demonstrated that anionic nanoliposomes as a carrier adjuvant could strongly induce a humoral immune response against displayed PCSK9 antigens. It was found that nanoliposomes displaying PCSK9-mimicking peptide linked to tetanus peptide can elicit long-term, strong, safe and specific antibodies against PCSK9 in BALB/c mice [22]. Our further study on C57BL/6 mice with severe atherosclerosis indicated that this vaccine setting, in a therapeutic approach, could generate functional and specific antibodies against PCSK9, which was associated with a long-lasting therapeutic effect on hypercholesterolemia and atherosclerosis [23]. Since the atherosclerotic plaque builds up gradually in the artery wall in which fat accumulation (fatty streaks) begins in early childhood and clinically relevant lesions become evident in middle-aged adults [24] and eventually, several years later, clinical manifestations become evident [25], preventing atheroma formation at its very initial stages can be an effective strategy to improve outcomes in individuals at risk of developing atherosclerosis [26]. To this end, the present study aimed to test the preventive efficacy of nanoliposome-displayed PCSK9 as an alternative peptide vaccine for PCSK9 inhibition against dyslipidemia and atherosclerosis in a C57BL/6 mouse model of severe atherosclerosis.

## Material and methods

### Preparation of nanoliposomal anti-PCSK9 vaccine

The lipid-film hydration approach was employed to manufacture liposome nanoparticles formulated

**Table I.** Sequence of the immunogenic peptides used in the present study

Peptide name	Sequence	Immunogenicity
PCSK9	S-I-P-W-N-L-E-R-I-T-P-V-R	B cell epitope
Tetanus	A-Q-Y-I-K-A-N-S-K-F-I-G-I-T-E-L	T cell epitope
IFPT	* <b>CGGGSIPWNLERITPVRKKAQYIKANSKFIGITEL</b>	

\*The bold amino acid codes exhibit linker sequences. IFPT – immunogenic fused PCSK9-tetanus.

with 1,2-dimyristoyl-sn-glycero-3-phosphorylglycerol (DMPG), 1,2-dimyristoyl-sn-glycero-3-phosphorylcholine (DMPC), and cholesterol (Chol). To construct vaccine formulation, an immunogenic peptide construct termed immunogenic fused PCSK9-tetanus (IFPT) (Table I) was linked to the nanoliposome surface using DSPE-PEG-maleimide (1,2-distearoyl-sn-glycero-3-phosphoethanolamine-*N*-[maleimide(PEG)-2000]) lipid, and then adsorbed onto Alum adjuvant [further details are described in the previous publication (22)].

#### Vaccination and atherosclerosis development

Thirty-five male C57BL/6 mice (6–8 weeks old, weighing 20 ±3 g) were obtained from the Laboratory Animal Research Center of the Pasteur Institute of Tehran, Iran. All animal treatments were performed in the basis of the Animal Welfare guidelines recommended by the European Parliament [27] and Institutional Ethics Committee and Research Advisory Committee of the Mashhad University of Medical Sciences, Mashhad, Iran. The animals were housed in an air-conditioned room at a constant temperature of 22 ±2°C with 12 : 12 h light/dark cycle and fed a standard mouse diet and water *ad libitum*. At the end of the study, all animals were sacrificed by sodium thiopental (30 mg/kg), intravenously [28, 29].

The mice were randomly sorted into five groups: 1) IFPT vaccine group, 2) IFPT plus Alhydrogel (IFPTA<sup>+</sup>) vaccine group, 3) nanoliposomal IFPT (L-IFPT) vaccine group, 4) L-IFPTA<sup>+</sup> vaccine group, 5) free-nanoliposome (negative control) group. All groups were subcutaneously vaccinated four times with the prepared vaccine formulations (200 µl in both the right and left flanks), once every 2 weeks. To develop atherosclerosis, the vaccinated and control mice were fed an atherogenic chow diet (containing 20% fat, 1.5% cholesterol, and 0.5% cholic acid) and concurrently treated every third day with intraperitoneal administration of poloxamer 407 (0.5 g/kg) for 16 weeks. This model has been shown to produce human-like pathological features of atherosclerosis in mice [30]. To measure titers of anti-PCSK9 antibody and plasma levels of lipid parameters, venous blood was taken at bi-weekly intervals throughout vaccination and post-vaccination periods for 16 weeks.

#### Analysis of vaccination effects

To measure the titers of anti-PCSK9 antibodies, plasma samples were isolated and assessed by ELISA technique. Concentrations of plasma PCSK9 in hypercholesterolemic vaccinated C57BL/6 mice were determined by a PCSK9 ELISA kit (CircuLex, Cy-8078, MBL, Woburn, MA) according to the manufacturer's manual. To verify targeting of mouse PCSK9 by vaccine-generated antibodies, PCSK9 binding to plasma antibodies was evaluated using the modified protocol of the PCSK9 ELISA kit [22]. CircuLex PCSK9-LDLR *in vitro* binding assay kit (CircuLex, Cy-8150, MBL, Woburn, MA) was utilized to determine the ability of plasma antibodies to suppress the PCSK9-LDLR interaction. Total cholesterol (TC), direct LDL-C, TG (triglyceride), and direct HDL-C (high-density lipoprotein cholesterol) were assessed using commercial kits. To measure expression of liver LDLR protein in vaccinated mice, Western blot analysis and immunohistochemical staining were carried out. The flow cytometry analysis was employed to determine the population of proinflammatory Th1 and anti-inflammatory Th2 cells, CD4<sup>+</sup> cells and their intracellular IFN-γ and IL-4 cytokines. At the end of the study, size and severity of atherosclerotic lesions were assessed in the aortic arch of vaccinated mice fed on an atherogenic diet, using hematoxylin and eosin (H&E) staining. The severity grade of lesions was classified between 1–4 [31]. Further details are described in the previous publication [23].

#### Statistical analysis

Unpaired 2-tailed Student's *t*-test and one-way ANOVA followed by Tukey's *post-hoc* test were carried out to define the significance of the difference among groups (GraphPad Prism Software, version 7, San Diego, CA). Data were reported as mean ± SD and lower-upper 95% confidence interval (CI) of mean, or median (interquartile range) for normally and non-normally distributed data, respectively. Data with *p* < 0.05 were regarded as statistically significant.

#### Results

##### Characterization of nanoliposomes

Characteristics of the free and IFPT-linked nanoliposome formulations are listed in Table II. The size

**Table II.** Physical properties of nanoliposomal formulations

Formulation	Z-average [nm] mean ± SD (n = 3)	Zeta potential [mV] mean ± SD (n = 3)	PDI mean ± SD (n = 3)
Empty nanoliposome	125 ±4.5	-43.9 ±3.8	0.01 ±0.001
IFPT-linked nanoliposome	155 ±5.5	-35.7 ±4.2	0.02 ±0.008

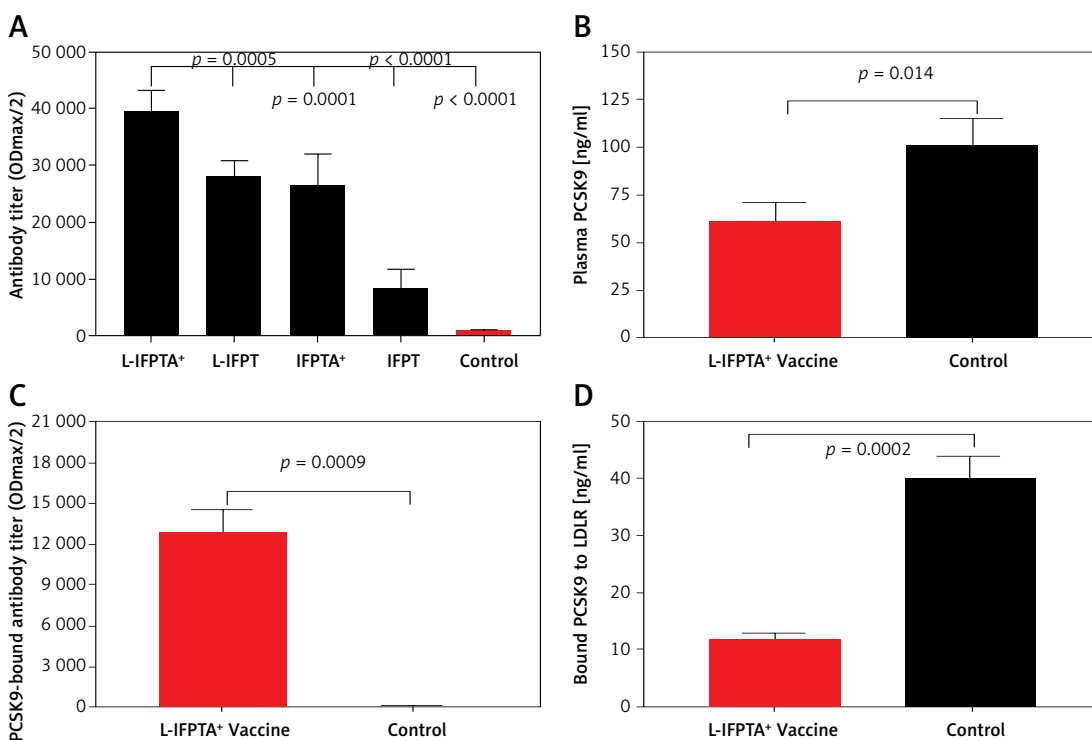
PDI – polydispersity index.

range of liposome nanoparticles was 120 nm to 155 nm in diameter, and the polydispersity index was < 0.1, indicating nanoparticles with high homogeneity. A negative zeta potential was also identified on the surface of the prepared nanoliposomes.

### Nanoliposomal vaccine provokes antibody against peptide PCSK9 in C57BL/6 mice

Among vaccine formulations, nanoliposomal vaccine displaying IFPT peptide formulated with Alhydrogel (L-IFPTA<sup>+</sup> vaccine) could induce the greatest IgG titers against PCSK9 peptide in the vaccinated mice. Two weeks after the last vaccination (week 8), hypercholesterolemic mice immunized with L-IFPTA<sup>+</sup> formulation indicated 1.41

(*p* = 0.0005), 1.5 (*p* = 0.0001), and 4.7 (*p* < 0.0001) fold higher serum antibody titers than mice immunized with the L-IFPT, IFPTA<sup>+</sup>, IFPT formulations, respectively (Figure 1 A). Notably, mice immunized by vaccines formulated with liposome nanoparticles showed higher titers of PCSK9 antibody than vaccine formulations without nanoliposome including IFPTA<sup>+</sup> and IFPT. In addition, as demonstrated by a long-term study, PCSK9 antibody in the L-IFPTA<sup>+</sup> vaccine group achieved peak titers at week 8 and stayed constant until week 12, then exhibited a decreasing trend (Figure 2 A). In conclusion, the findings reveal effective adjuvant potential of liposome nano-carriers to provoke an antibody response against PCSK9 peptide antigen.



**Figure 1.** Efficacy of antiPCSK9 vaccines in C57BL/6 mice, 2 weeks after the last vaccination. **A** – Antibody titers (ODmax/2) against PCSK9 peptide in the plasma of mice vaccinated with various vaccine formulations or mice inoculated with free nanoliposomes as a control. **B** – Plasma levels of PCSK9 in vaccinated and control groups were 61.37 ±5.53 ng/ml and 101.5 ±8.04 ng/ml, respectively. **C** – Direct detection of antibodies bound to plasma PCSK9 (target engagement) in blood samples from vaccinated and control mice. Increased ODmax/2 is indicative for vaccine-generated PCSK9 antibodies directly binding to PCSK9. **D** – *In vitro* PCSK9/LDLR binding assay. Vaccine-generated antiPCSK9 antibodies inhibit PCSK9-LDLR interaction. Plasma sample of L-IFPTA<sup>+</sup> vaccine group decreased PCSK9 binding to LDLR by 69.86%, compared with plasma sample of the control group. Values are means ± SD, *n* = 3 replicates of the pooled samples of 7 mice per group. Significance compared to control values was analyzed by unpaired 2-tailed Student’s *t*-test. Statistical differences at *p*-values less than 0.05 were considered to be significant

IFPT – immunogenic fused PCSK9-tetanus, L-IFPT – nanoliposomal IFPT, IFPTA<sup>+</sup> – IFPT plus Alhydrogel, L-IFPTA<sup>+</sup> – nanoliposomal IFPT plus Alhydrogel.

### Impact of vaccine-provoked PCSK9 antibodies on plasma PCSK9 levels

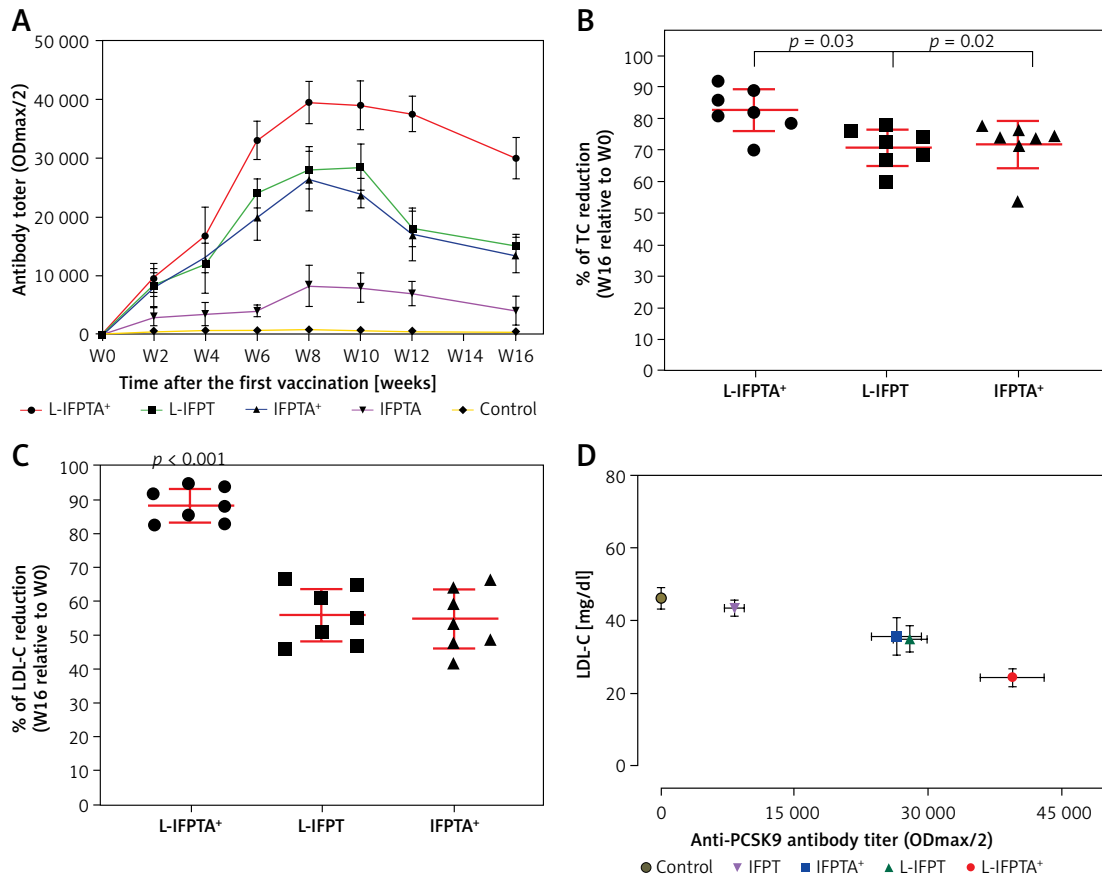
Vaccine-provoked PCSK9 antibodies in hypercholesterolemic C57BL/6 mice were detected to reduce plasma levels of PCSK9 through specific targeting. Blood concentrations of PCSK9 in the L-IFPTA<sup>+</sup> vaccine and control group were found to be 61.37 ±5.53 ng/ml and 101.5 ±8.04 ng/ml, respectively (Figure 1 B). The level of plasma PCSK9 was significantly lower by 40% (-40.13 ±9.758, *p* = 0.014) in vaccinated mice than control mice. To verify specific targeting, an ELISA-based approach was employed [22] to detect plasma PCSK9-bound antibodies. As revealed in Figure 1 C, the L-IFPTA<sup>+</sup> vaccine group exhibited high titers of PCSK9-bound antibody, verifying direct binding of vaccine-induced antibodies to plasma PCSK9. Overall, L-IFPTA<sup>+</sup> vaccine can provoke specific antibodies that directly target and eliminate circulating PCSK9 from the bloodstream.

### Impact of vaccine-provoked PCSK9 antibodies on PCSK9 function

To determine whether the PCSK9 antibodies can suppress PCSK9 function, the *in vitro* binding of PCSK9 and LDLR in the presence of plasma PCSK9 antibodies was assayed. It was found that a plasma sample of the L-IFPTA<sup>+</sup> vaccine group could significantly reduce *in vitro* PCSK9-LDLR binding by 69.86%, in comparison with a plasma sample of the control group (Figure 1 D). Overall, the L-IFPTA<sup>+</sup> vaccine induced generation of PCSK9 antibodies in hypercholesterolemic C57BL/6 mice, which can suppress PCSK9-LDLR interaction and thereby inhibit PCSK9 function.

### Liposomal vaccines alleviate hypercholesterolemia in C57BL/6 mice

The liposomal vaccine showed a protective effect against hypercholesterolemia in C57BL/6



**Figure 2.** Long-term efficacy of liposomal vaccines in hypercholesterolemic mice. **A** – Long-term evaluation of antibody titers over 16 weeks after prime vaccination against PCSK9 peptide (ODmax/2). Data are shown as means ± SD (*n* = 7 per group). Long-term changes of plasma TC (**B**) and LDL-C (**C**) in the vaccinated hypercholesterolemic C57BL/6 mice, at week 16 compared to week 0 (pre-vaccination) adjusted to control group. TC (**B**) and LDL-C (**C**) levels were reduced in the L-IFPTA<sup>+</sup>, L-IFPT, and IFPTA<sup>+</sup> vaccine group 8 weeks after the last vaccination. Data are shown as means with 95% confidence intervals (*n* = 7 per group). **D** – Correlation plot of circulating LDL-C levels with PCSK9 antibody titers evaluated 2 weeks after the last vaccination (week 8) with L-IFPTA<sup>+</sup>, L-IFPT, and IFPTA<sup>+</sup> vaccine and control liposome. A significant negative correlation was identified among LDL-C and the PCSK9 antibodies in the vaccinated and in control mice (*r* = -0.94, *p* = 0.008). One-way ANOVA followed by Tukey's *post-hoc* test was used to determine the significance of the difference among groups. Statistical differences at *p*-values less than 0.05 were considered to be significant

mice. The L-IFPTA<sup>+</sup> vaccine resulted in an increasing trend of plasma TC and LDL-C in hypercholesterolemic C57BL/6 mice, at week 8 (Figure 3). TC was lower in hypercholesterolemic C57BL/6 mice vaccinated with L-IFPTA<sup>+</sup> ( $-38.13 \pm 3.8\%$ , 95% CI: 34–41%,  $p = 0.006$ ), L-IFPT ( $-23 \pm 4.1\%$ , 95% CI: 19–27%,  $p = 0.027$ ), and IFPTA<sup>+</sup> ( $-19.12 \pm 3\%$ , 95% CI: 16–22%,  $p = 0.038$ ) than in the control group (Figures 3 A, E). LDL-C was reduced in hypercholesterolemic mice vaccinated with L-IFPTA<sup>+</sup> ( $-57 \pm 7.7\%$ , 95% CI: 50–64%,  $p = 0.0003$ ), L-IFPT ( $-41.67 \pm 4.2\%$ , 95% CI: 37–45%,  $p = 0.03$ ), and IFPTA<sup>+</sup> ( $-36.11 \pm 5\%$ , 95% CI: 31–40%,  $p = 0.02$ ) than in the control group (Figures 3 B, F). Plasma HDL-C and TG levels did not differ between control and vaccinated groups (Figures 3 C, D).

### Long-lasting impact of liposomal vaccines in hypercholesterolemic mice

A long-term follow-up was performed to evaluate the durability of antibody response against PCSK9 as well as the alterations in plasma cholesterol levels in hypercholesterolemic vaccinated mice. Analysis of antibody titers during 16 weeks after immunization with L-IFPTA<sup>+</sup>, L-IFPT, and IFPTA<sup>+</sup> vaccine formulations revealed that a long-lasting humoral immune response was persistently induced against PCSK9 peptide (Figure 2 A), which was correlated with a significant reduction of TC in L-IFPTA<sup>+</sup>, L-IFPT, and IFPTA<sup>+</sup> vaccine groups by up to  $-82.5 \pm 7.3\%$  (95% CI: 75–89%,  $p = 0.002$ ),  $-70.54 \pm 6.2\%$  (95% CI: 65–76%,  $p = 0.013$ ),  $72.02 \pm 8.7\%$  (95% CI: 64–79%,  $p = 0.004$ ), respectively (Figure 2 B), and LDL-C up to  $-88.14 \pm 5.6\%$  (95% CI: 84–93%,  $p = 0.002$ ),  $-55.92 \pm 8.3\%$ , (95% CI: 48–63%,  $p = 0.003$ ) and  $54.81 \pm 9.3\%$  (95% CI: 46–63%,  $p = 0.003$ ), respectively (Figure 2 C), compared to the pre-vaccination time point (W0) adjusted to the control group.

### Inverse association of PCSK9 antibody titers and LDL-C levels in vaccinated mice

The statistical analysis revealed a significant negative association between plasma concentrations of LDL-C and titers of PCSK9 antibodies in the hypercholesterolemic vaccinated mice, in which LDL-C and PCSK9 antibodies were respectively at the lowest and highest levels. Therefore, the anti-PCSK9 vaccine formulations, to a higher extent than the L-IFPTA<sup>+</sup> vaccine, provoked high titers of PCSK9 antibodies that are directly associated with the decrease in plasma concentrations of LDL-C.

### Liposomal vaccines increase the hepatic LDLR

Western blot and immunohistochemical (IHC) analyses were performed to evaluate the impact

of liposomal anti-PCSK9 vaccine on the expression and cellular distribution of the hepatic LDLR protein in hypercholesterolemic C57BL/6 mice (Figure 4). The hepatic LDLR protein was detected by Western blotting followed by quantification using ImageJ (Figure 4 A). When normalized to  $\beta$ -actin, the protein expression of hepatic LDLR in hypercholesterolemic mice vaccinated with L-PTA<sup>+</sup>, L-IFPT, and IFPTA<sup>+</sup> was found to be  $2.93 \pm 0.32$  (95% CI: 2.1–3.7,  $p = 0.0001$ ),  $1.55 \pm 0.4$  (95% CI: 0.4–2.4,  $p = 0.03$ ), and  $1.39 \pm 0.4$  (95% CI: 0.2–2.2,  $p = 0.06$ ) fold greater than in the negative control mice (Figure 4 B). As revealed by IHC analyses of the same liver samples, approximately similar staining of the cell membrane and cytoplasm was detected in the negative control group (Figure 4 C), while samples from vaccinated mice showed considerably increased staining at the cell membrane (Figure 4 D). Hence, nanoliposomal vaccines can efficiently increase the cell surface levels of hepatic LDLR in vaccinated hypercholesterolemic mice.

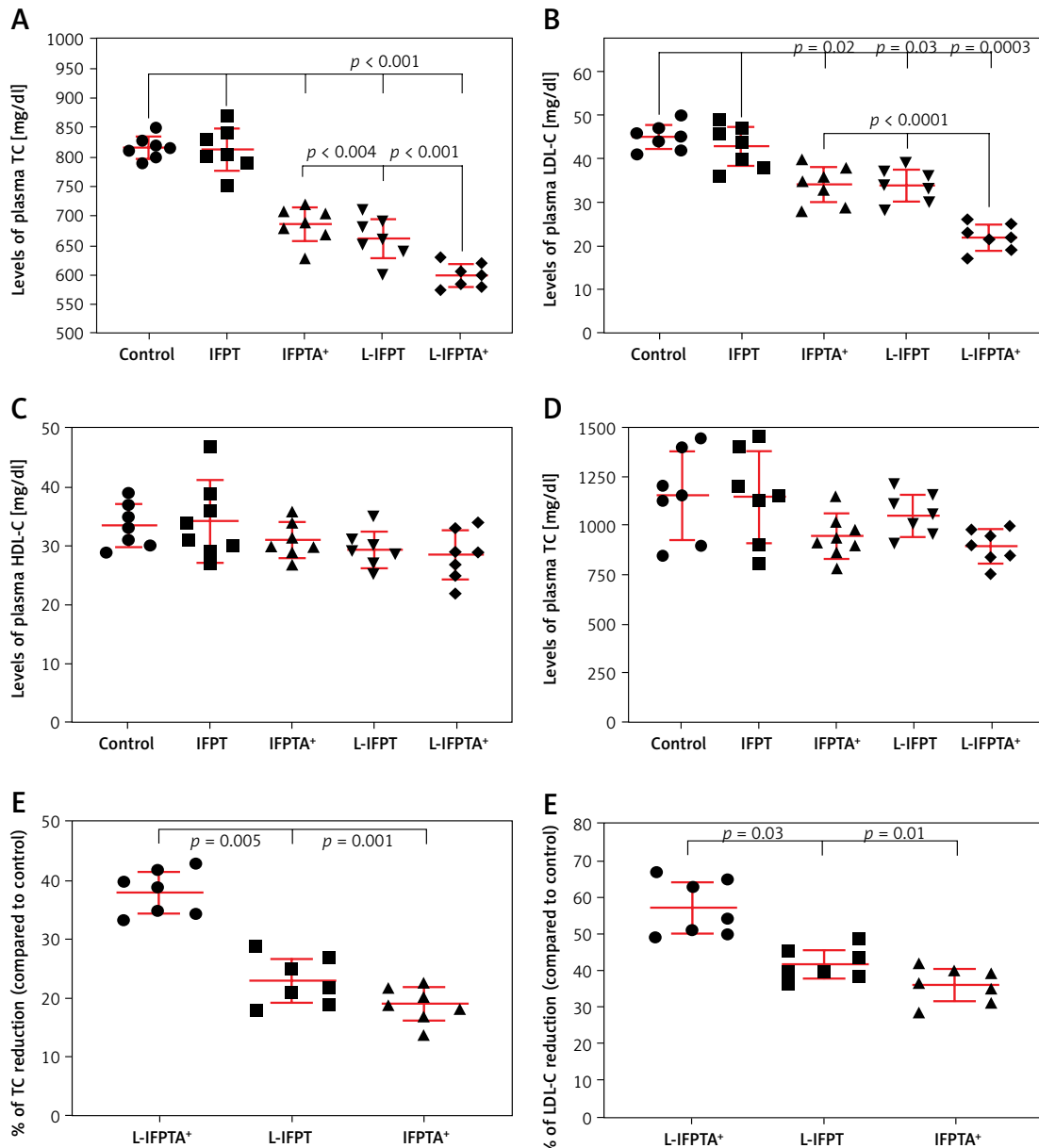
### Liposomal vaccine prevents aortic atherosclerotic lesions

To evaluate the protective impact of the liposomal anti-PCSK9 vaccine on atherosclerosis lesion progression, lesion size, intima-to-media thickness (IMT), and lesion grade were assessed in the aortic arc upon a 16-week atherogenic diet (Figure 5). Mice immunized with the L-IFPTA<sup>+</sup> vaccine exhibited a significantly decreased lesion size (24.25%,  $p = 0.002$ ) and IMT (30.2%,  $p = 0.007$ ), compared with the negative controls (Figures 5 A, B). Moderate lesions (grade II) were observed in 57.2% and 28.5% of L-IFPTA<sup>+</sup> and control mice, respectively; however, more severe lesions (grade III–IV) were only detected in control mice (71.5%), while 42.85% of L-IFPTA<sup>+</sup> mice demonstrated grade I lesions (Figure 5 C). Histological images of H&E-stained aortic arch lesions of vaccinated and negative control mice are presented in Figure 5 D. A significant and strong positive association was found between the size of aortic atherosclerotic lesions and plasma levels of LDL-C ( $r = 0.83$ ,  $p = 0.001$ ) and PCSK9 protein ( $r = 0.71$ ,  $p = 0.0024$ ).

### Liposomal vaccine exerts anti-inflammatory impact in hypercholesterolemic mice

Flow cytometry analysis of splenocytes indicated that the L-IFPTA<sup>+</sup> vaccine was immunologically safe and increased anti-inflammatory response in hypercholesterolemic mice.

The L-IFPTA<sup>+</sup> vaccine could increase the anti-inflammatory Th2 cell population (Figures 6 A1,



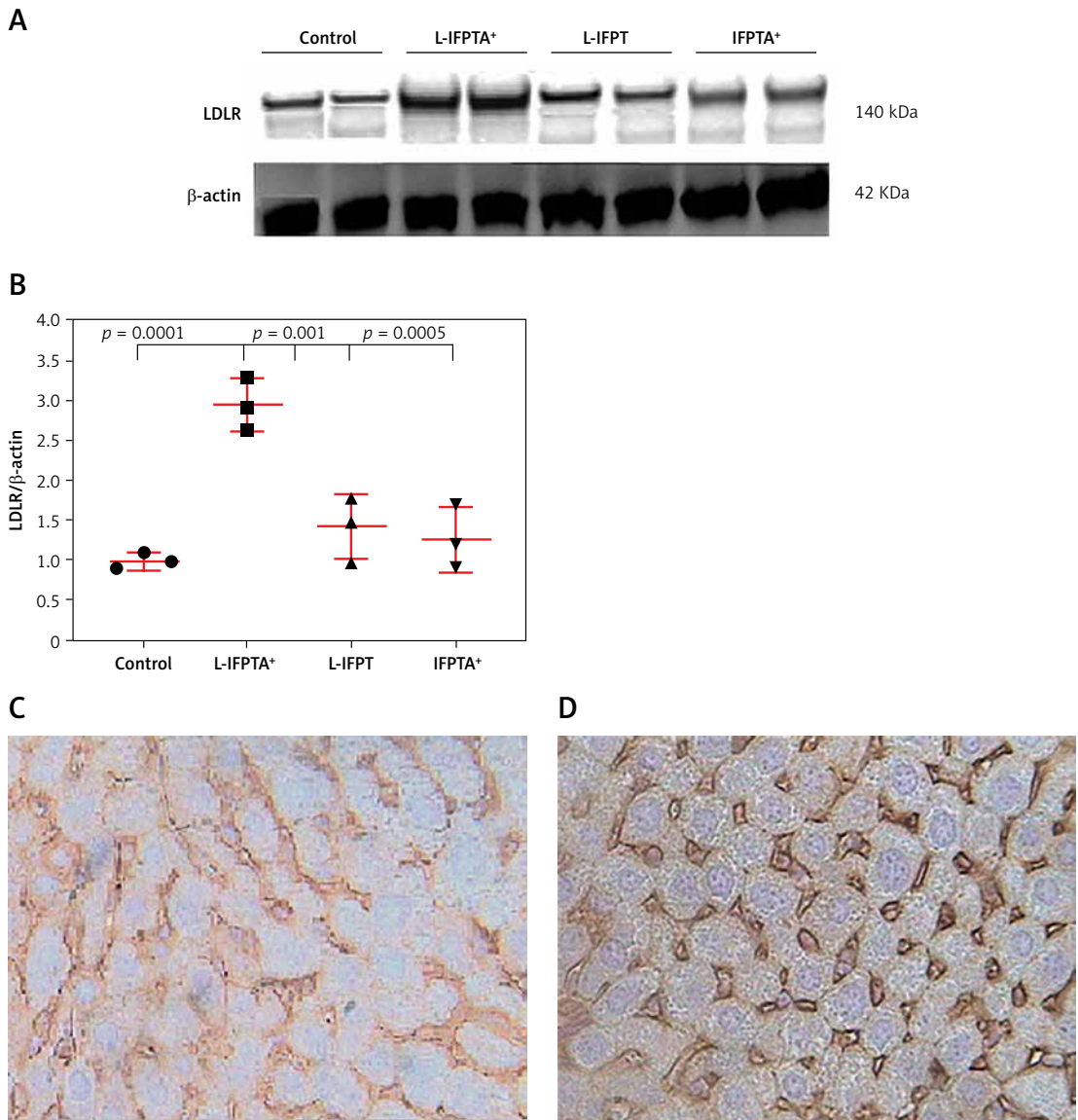
**Figure 3.** Lipid-lowering effect of anti-PCSK9 vaccine formulations. Levels of plasma TC (A), LDL-C (B), HDL-C (C), and TG (D) in hypercholesterolemic C57BL/6 mice, 2 weeks after the fourth vaccination (week 8) with various anti-PCSK9 vaccine formulations. Compared with the control group, TC (E) and LDL-C (F) levels were significantly decreased in the L-IFPTA+, L-IFPT, and IFPTA+ vaccine group 2 weeks after the last vaccination. There was no significant difference of HDL-C and TG between the control and the vaccinated groups. Data are shown as means with 95% confidence interval ( $n = 7$  per group). One-way ANOVA followed by Tukey's *post-hoc* test was used to analyze the significance of the difference among groups. Statistical differences at  $p$ -values less than 0.05 were considered to be significant

HDL-C – high-density lipoprotein cholesterol, LDL-C – low-density lipoprotein cholesterol, TC – total cholesterol, TG – triglyceride.

A3) and corresponding IL-4 cytokine in vaccinated mice (Figures 6 B1, B3), whereas the baseline frequency of the pro-inflammatory Th1 cell population (Figures 6 A2, A4) and INF- $\gamma$  cytokine level (Figures 6 B2, B4) were not significantly changed. Compared with the control naïve mice, the population of Th2 cells (Figure 6 A3) and production of IL-4 cytokine (Figure 6 B3) were 1.76- and 4.9-fold higher in the L-IFPTA+ vaccine group.

## Discussion

The current research demonstrated that active immunotherapy against PCSK9 using a nanoliposomal peptide-based vaccine termed L-IFPTA+ could protect against hypercholesterolemia and atherosclerosis in mice immunized with a prophylactic route. Nanoliposomal anti-PCSK9 vaccine promoted functional and specific antibodies that targeted



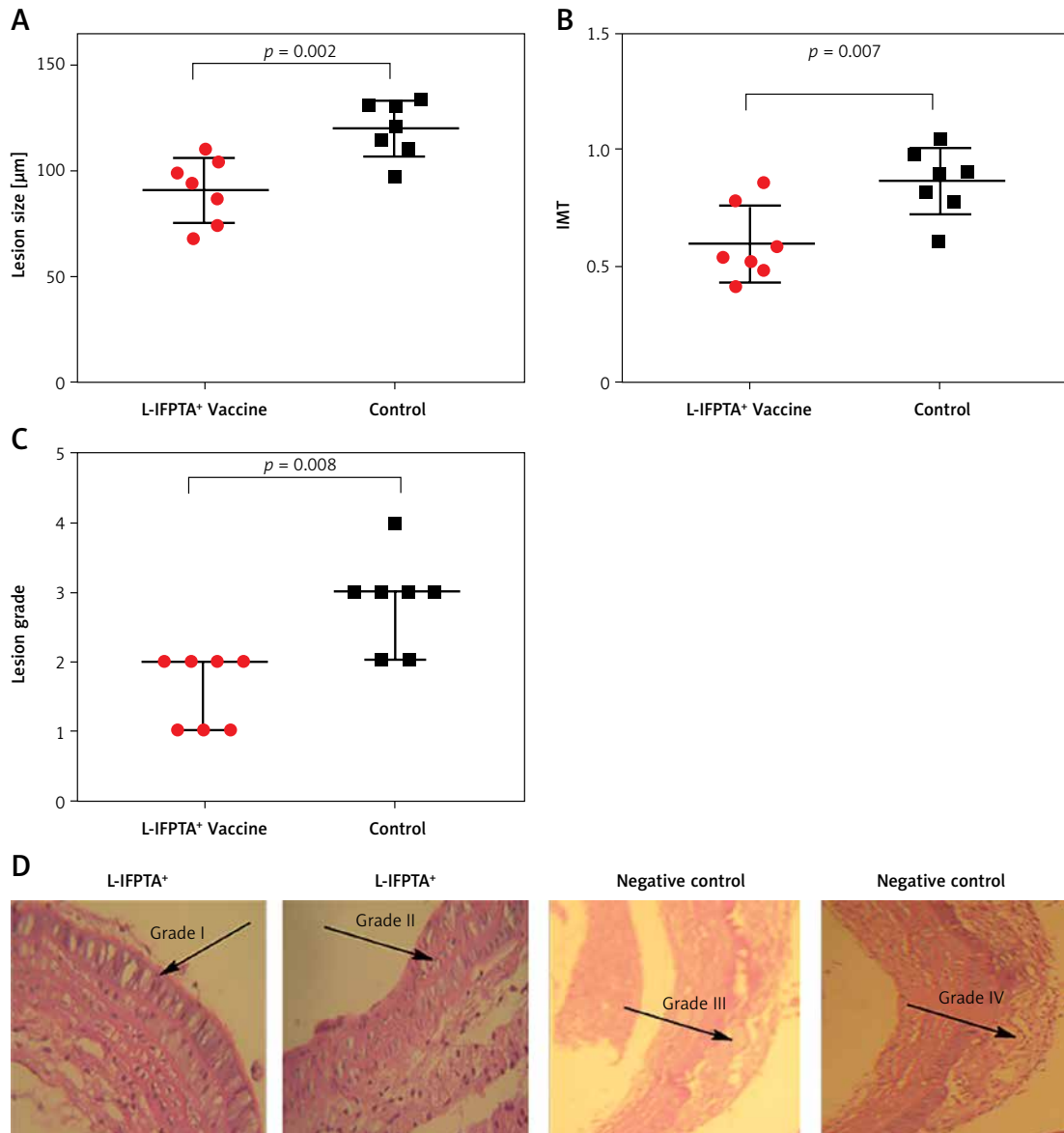
**Figure 4.** Expression (A, B) and cellular distribution (C, D) of LDLR in livers of vaccinated mice. Liver extracts from mice vaccinated with L-IFPTA<sup>+</sup>, L-IFPT, and IFPTA<sup>+</sup>, or from unvaccinated (control) mice were separated by SDS-PAGE and blotted onto a nitro-cellulose filter. Following incubation with HRP-conjugated anti-LDLR or anti-β actin antibody, the bands corresponding to LDLR or β-actin proteins were measured by ImageJ software. The LDLR bands were normalized on the corresponding β-actin band (A). Compared to the control group, the protein expression of LDLR in the liver was significantly increased in the L-IFPTA<sup>+</sup> vaccine group. Data are expressed as means with 95% confidence interval (*n* = 3 per group). One-way ANOVA followed by Tukey's *post-hoc* test was used to determine the significance of the difference among groups. Statistical differences at *p*-values less than 0.05 were considered to be significant (B). Cellular distribution of LDLR in immunized mice (C, D). Immuno-histochemical analysis was carried out on liver samples isolated from the control (C) or vaccinated (D) mice using rabbit anti-LDLR antibodies detected by anti-rabbit IgG HRP-conjugated antibodies

circulating PCSK9 and suppressed its interaction with LDLR, resulting in a significant elevation of liver LDLR and subsequent reduction of TC and LDL-C. Additionally, significant reductions of inflammatory cells (Th1) and IFN-γ cytokine as well as significant elevations of anti-inflammatory cells (Th2) and IL-4 cytokine were observed, which were parallel to the reduction of aortic atherosclerosis lesion size in hypercholesterolemic C57BL/6 mice.

PCSK9 has a pivotal role in LDL-C metabolism through promoting degradation of liver LDLR,

which is the main route for clearance of circulating LDL-C. PCSK9 inhibition is considered to be the strongest therapeutic approach for reducing plasma LDL-C and atherosclerotic events. Evolocumab and alirocumab are two FDA-approved PCSK9 mAbs that were recommended by the 2016 ESC/EAS Guideline as second-line treatment for the management of dyslipidemias in hypercholesterolemic patients with an inadequate response to statin therapy [32]. Nevertheless, mAbs possess intrinsic disadvantages including



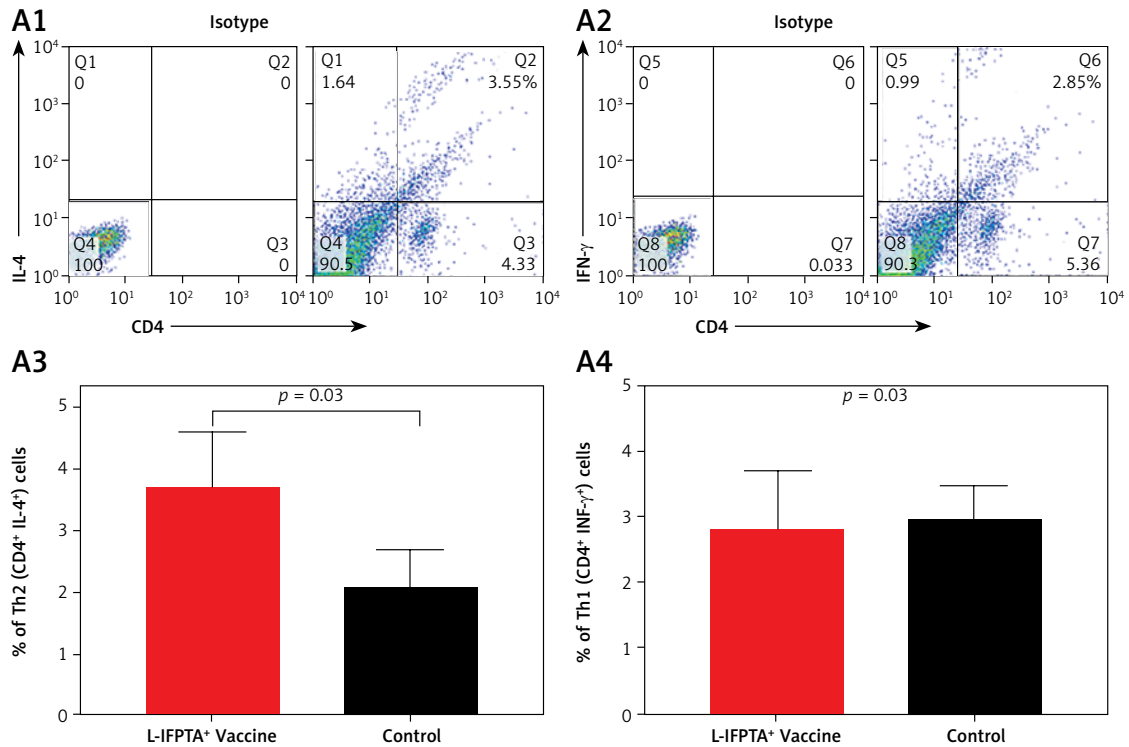


**Figure 5.** Inhibitory effect of L-IFPTA<sup>+</sup> vaccine on development of atherosclerotic lesion in aortic arc area of C57BL/1 mice model of diet induced-atherosclerosis. 16 weeks after prime vaccination of mice on severe atherogenic diet lesion development was determined using hematoxylin and eosin (H&E) staining. **A** – Lesion size in vaccinated and control mice was 90.86 ± 5.849 µm and 120 ± 4.995 µm, respectively. Lesion size was found to be 24.25% (–29.14 ± 7.692 µm,  $p = 0.002$ ) lower in vaccinated mice than control mice. Data are expressed as group means ± SD ( $n = 7$  mice per group). Significance compared to control values was analyzed by unpaired 2-tailed Student's  $t$ -test. **B** – Intima-to-media thickness (IMT) in vaccinated and control mice was 0.5978 ± 0.06271 and 0.8634 ± 0.05393, respectively. IMT in vaccine group was significantly decreased by 30.2% (0.2656 ± 0.08271 µm,  $p = 0.007$ ) compared with control group. Data are expressed as group means ± SD ( $n = 7$  mice per group). Significance compared to control values was analyzed by unpaired 2-tailed Student's  $t$ -test. **C** – Lesion grade in vaccinated and control mice was 2 [1-2] and 3 [2-3], respectively. Data are expressed as group medians [25<sup>th</sup> percentile – 75<sup>th</sup> percentile] ( $n = 7$  mice per group). Significance compared to control values was analyzed by Mann-Whitney  $U$  test. Statistical differences at  $p$ -values less than 0.05 were considered to be significant. **D** – Representative images of H&E-stained grade I, II, III, and IV lesions of aortic arc

high-dose and frequent administration [33], possible induction of host anti-mAb antibodies [34], and high cost [35], which restrict their clinical use for chronic diseases demanding long-term treatments. To overcome such drawbacks, active immunotherapy with long-lasting effects can be a cost-effective and safe alternative to mAb-

based therapy of hypercholesterolemia and atherosclerosis [34, 36].

In a previous study, we demonstrated that our liposomal peptide-based active immunization approach could strongly elicit a durable and safe PCSK9-specific antibody response in vaccinated BALB/c mice [22]. Accordingly, it was shown that

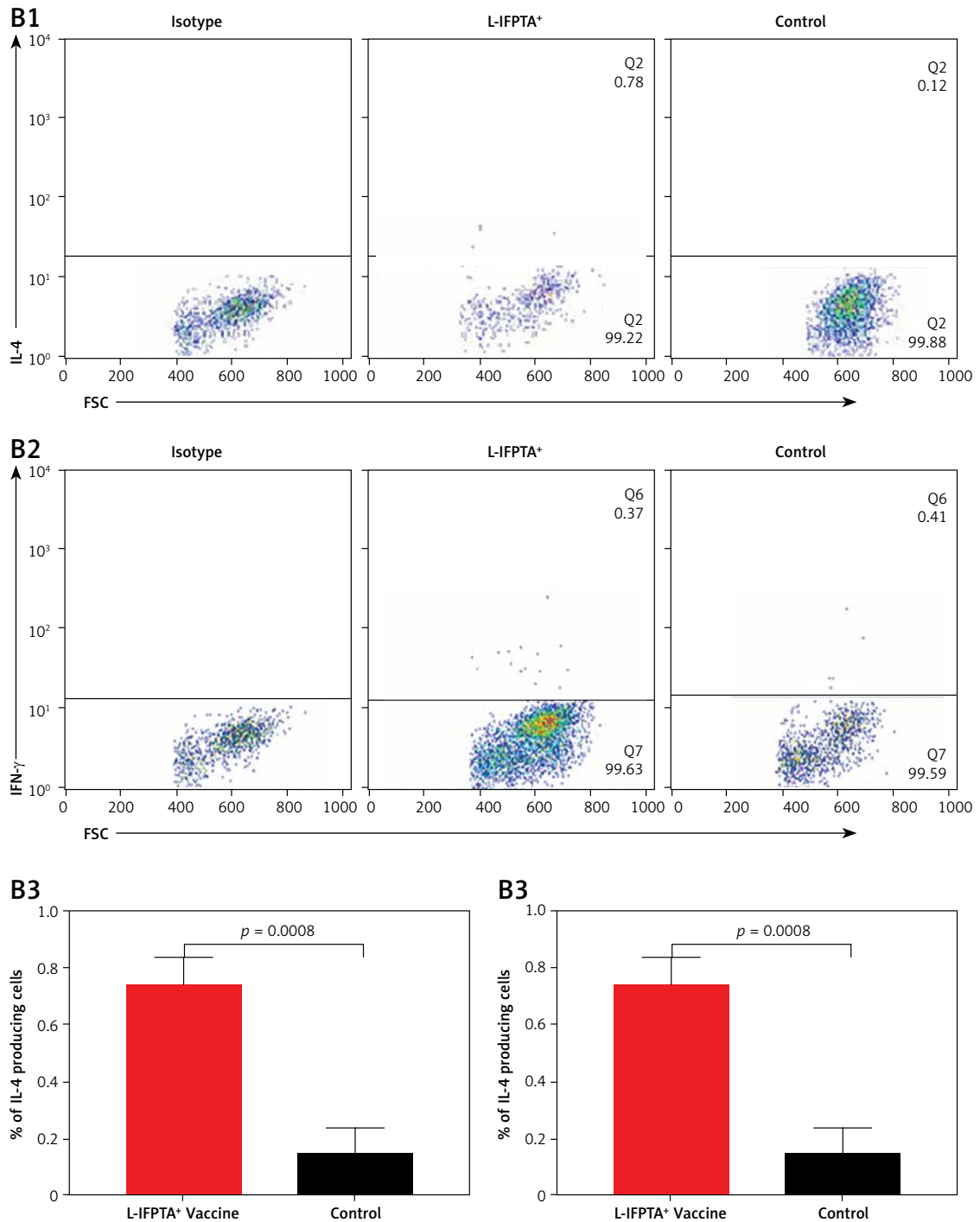


**Figure 6. A** – L-IFPTA<sup>+</sup> vaccine increase anti-inflammatory Th2 cells in hypercholesterolemic C57BL/6 mice. Representative scattergrams exhibit gating strategy and quadrant analysis for flow cytometry of Th2 (CD4<sup>+</sup> IFN- $\gamma$ <sup>+</sup> T cell) (A1) and Th1 (CD4<sup>+</sup> IL-4<sup>+</sup> T cell) (A2) population on the basis of isotype control. Splenic Th2 cell population in L-IFPTA<sup>+</sup> vaccine group were higher than in naïve control mice (A3), while there was no significant difference in Th1 cell population of vaccine and control group (A4)

the liposomal anti-PCSK9 vaccine could strongly decrease LDL-C and inhibit progression of atherosclerotic lesion in C57BL/6 mice on a harsh atherogenic regimen [23]. Vaccine-based PCSK9 suppression has also been evaluated by others using different approaches. AFFIRIS developed an AFFITOPE-based PCSK9 active immunization approach [14] that decreased TC by 53%, which was correlated with decreased size and severity of atherosclerosis plaque in vaccinated compared with control mice [15]. Likewise, Fattori *et al.* studied a combination of human recombinant PCSK9 and DNA oligonucleotide as an adjuvant, and observed an approximately 40% decrease of LDL-C in vaccinated mice [16]. Pan *et al.* developed bacteriophage virus-like particles, on top of which PCSK9 peptides were displayed at a high valency. This human PCSK9 vaccine reduced TC levels by ~20% compared with the control [17].

We are the first to present the liposomal anti-PCSK9 vaccine L-IFPTA<sup>+</sup> that exhibits prophylactic effects against hypercholesterolemia and atheroma formation in a C57BL/6 mouse model of severe atherosclerosis. The L-IFPTA<sup>+</sup> vaccine elicits high-titer specific antibodies by which plasma levels of PCSK9 in hypercholesterolemic mice were decreased by up to 40%, compared with the control mice. Target engagement analysis

*in vivo* revealed that L-IFPTA<sup>+</sup>-induced antibodies bound specifically to plasma PCSK9 in vaccinated mice. *In vitro* analysis of PCSK9-LDLR interaction demonstrated that the L-IFPTA<sup>+</sup> vaccine generated functional PCSK9 antibodies that inhibited interaction between PCSK9 and LDLR. It is well documented that inhibition of the PCSK9-LDLR interaction causes up-regulation of LDLR on the surface of hepatocytes, thereby enhancing the uptake of plasma cholesterol from the bloodstream [16, 37]. In line with this evidence, our approach inhibited the PCSK9-LDLR interaction, which consequently resulted in LDLR up-regulation and subsequent reduction of plasma cholesterol, particularly LDL-C. The L-IFPTA<sup>+</sup> vaccine inhibited the increase of plasma TC and LDL-C as well as progression of atherosclerotic lesion in C57BL/6 mice on a severe atherogenic diet plus poloxamer 407 injection. Two weeks after the fourth vaccination (week 8), the mean PCSK9 antibody titers reached the highest levels in hypercholesterolemic mice and resulted in reductions of TC and LDL-C by up to –38.13% and –55%, respectively. The results are supported by statistical analysis that showed a significant inverse correlation between plasma levels of LDL-C and PCSK9 antibody titers. Long-term monitoring (16 weeks) showed that during 10 weeks after the last vaccination (weeks 6–16) the antibody



**Figure 6.** Cont. **B** – L-IFPTA<sup>+</sup> vaccine increased anti-inflammatory IL-4 cytokine in hypercholesterolemic C57BL/6 mice. Representative scattergrams exhibit gating strategy and quadrant analysis for flow cytometry of IL-4 (**B1**) and IFN- $\gamma$ <sup>+</sup> (**B2**) producing splenocytes on the basis of isotype control. Expression of IL-4 in L-IFPTA<sup>+</sup> vaccine group was significantly higher than in naïve control mice (**B3**), while there was no significant difference of IFN- $\gamma$ <sup>+</sup> between vaccine and control groups (**B4**). Data are expressed as mean  $\pm$  SD of 7 mice in each group. Significance compared to control values was analyzed by unpaired 2-tailed Student's *t*-test. Statistical differences at *p*-values less than 0.05 were considered to be significant

titer in vaccinated mice remained significantly higher than that in the control group, which was accompanied by significant reductions in TC and LDL-C at these time points. Adjusting data to the control and comparing them to the onset time

point revealed that at the end of the study (during 16 weeks) TC was decreased by up to –82.5% and LDL-C up to –88.14% in vaccinated mice on a severe atherogenic diet, which was associated with significantly lower lesion size (–24.25%, *p* = 0.002)

and severity ( $-45\%$ ,  $p = 0.008$ ) in the aortic arch compared with the control.

An inhibitory effect of the PCSK9 vaccine on progression of atherosclerotic lesions was also observed in APOE\*3Leiden.CETP mice [15]; however, our study demonstrated the atheroprotective effect of L-IFPTA<sup>+</sup> in mice receiving a concomitant atherogenic diet plus poloxamer 407 injection to induce severe atherosclerosis. This model has repeatedly been shown to be reliable for studying atherosclerosis and its pathological features in mice [38]. Nevertheless, an important limitation for developing PCSK9 inhibitors is the lack of a reliable mouse model that can recapitulate high levels of LDL-C in humans. Indeed, in the mouse, unlike the human, a large amount of cholesterol is eliminated by HDL metabolism, and thus baseline levels of LDL-C are not sufficiently increased as those witnessed in human [38]. Therefore, this can reveal a possible reason for the moderately lower effectiveness of PCSK9 inhibitors when trialed in humans [14, 15]. For example, despite the efficient LDL-C lowering of an anti-PCSK9 vaccine in preclinical studies, its effect was less impressive, with a 13% LDL-C level reduction, when tested in a phase I clinical trial (Bauer *et al.*, Communication at ESC Congress, 2018 Munich). Notably, although in the present study we designed a preventive approach in which baseline level of LDL-C was low in hypercholesterolemic mice when vaccination was primed, in our previous study using the therapeutic approach, an elevated level of baseline LDL-C ( $\sim 200$  mg/dl) was evidenced [23], suggesting that the hypercholesterolemic mouse model developed in this study may be a model near to human hypercholesterolemia and hence can provide a reliable model that extrapolates preclinical data of anti-PCSK9 inhibitors to humans. Thus, the preventive effect of the L-IFPTA<sup>+</sup> vaccine observed in the current study can introduce a promising strategy for effective prevention of increasing levels of LDL-C and atherosclerotic plaque progression in humans.

Inflammation is known to have an important role in atherosclerosis progression and complications. T lymphocytes, particularly subsets of CD4<sup>+</sup> helper T (Th) cells, have been found to play an important role in atherosclerosis, both in humans [39] and in animals [40]. These CD4-bearing Th cells are classified mainly into Th1 (CD4<sup>+</sup> IFN- $\gamma$ <sup>+</sup>) and Th2 (CD4<sup>+</sup> IL-4<sup>+</sup>) subtypes showing pro-atherogenic and anti-atherogenic roles, respectively [41–43]. The pro-inflammatory IFN- $\gamma$  cytokine can induce and worsen atherosclerosis progression via influencing lipid accumulation and foam cell formation in the vascular wall, and cellular composition of the plaque, such as inducing proliferation of SMCs, macrophage activation, autocrine

induction of Th1 cells, and suppressing proliferation of Th2 cells [44]. In contrast, IL-4 secreted by Th2 cells counteracts the IFN- $\gamma$  production [45]. Further, high levels of Th2 cells in the circulating blood were independently associated with a decrease in carotid plaque thickness, and a reduced risk of acute myocardial infarction in females [45]. In the present study, it was found that the population of anti-inflammatory Th2 cells as well as production of IL-4 cytokine were significantly increased by 1.76- and 4.9-fold, respectively, in splenic cells isolated from the L-IFPTA<sup>+</sup>-vaccinated mice compared with controls. This finding implies that the L-IFPTA<sup>+</sup> vaccine induced a safe immune response in hypercholesterolemic mice.

In conclusion, the L-IFPTA<sup>+</sup> vaccine induced safe, durable and functional PCSK9-specific antibodies in hypercholesterolemic C57BL/6 mice. This resulted in a long-term therapeutic, as reported previously [23], and protective effects against hypercholesterolemia and atherosclerosis. The present findings suggest the potential of the L-IFPTA<sup>+</sup> vaccine as a potentially effective alternative to mAb-based therapies for controlling elevated cholesterol levels and preventing cardiovascular disease.

### Conflict of interest

The authors declare no conflict of interest.

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